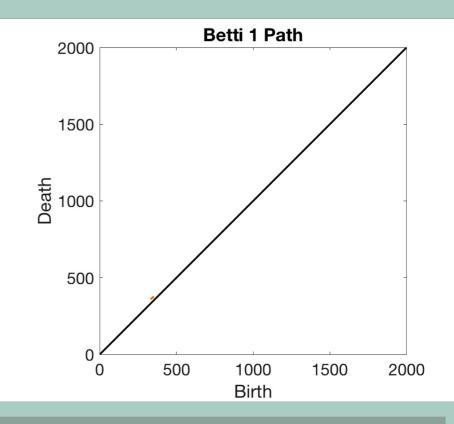
TOPOLOGICAL DATA ANALYSIS FOR BIOLOGICAL RING CHANNELS

Veronica Ciocanel OSU President's Postdoctoral Scholar Mathematical Biosciences Institute

The Ohio State University



SIAM Dynamical Systems Snowbird, UT May 21st 2019



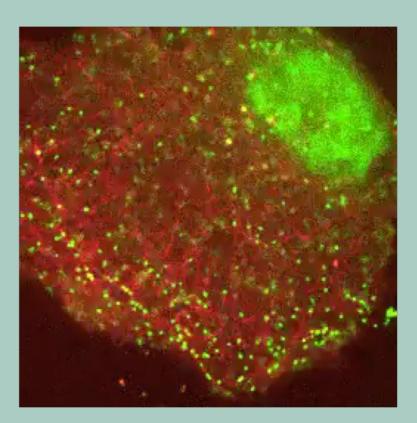
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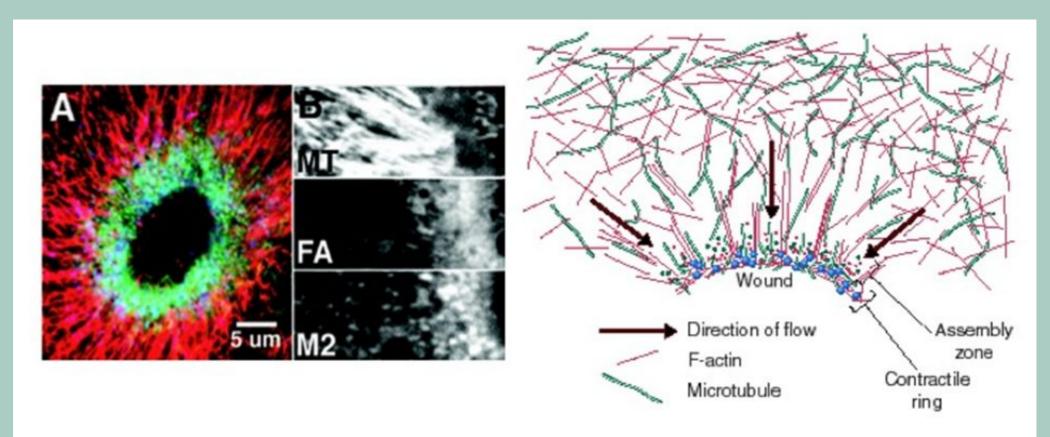


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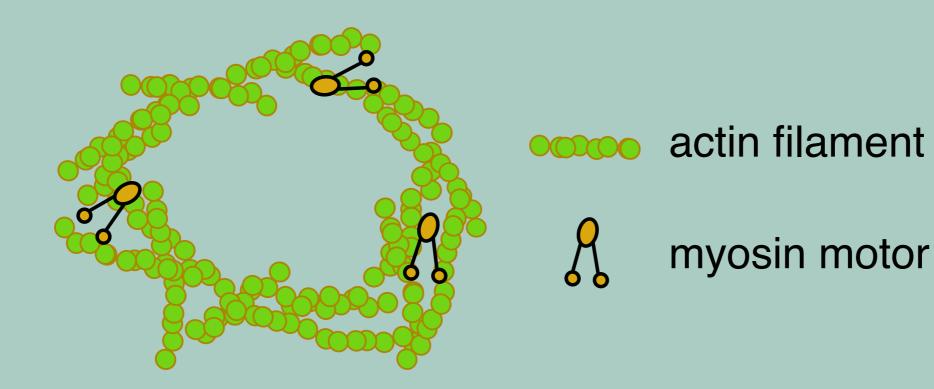
MOTIVATION

- Cells use many channels to communicate, each with different functions.
- Ring channels play critical roles in oogenesis, wound healing, and cell division.

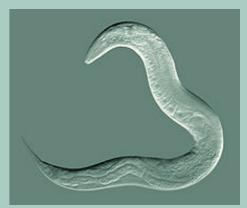


MOTIVATION

- Often, ring channels maintain precise diameters over a large time scale.
- Non-muscle myosin motor proteins are often "in charge" of creating appropriate constriction of the ring.



SPECIFIC MOTIVATION



Biophysical Journal



Antagonistic Behaviors of NMY-1 and NMY-2 Maintain Ring Channels in the *C. elegans* Gonad

Valerie C. Coffman,¹ Torah M. Kachur,² David B. Pilgrim,² and Adriana T. Dawes^{1,3,*} ¹Department of Molecular Genetics, The Ohio State University, Columbus, Ohio; ²Department of Biological Sciences, University of Alberta, Edmonton, Alberta Canada; and ³Department of Mathematics, The Ohio State University, Columbus, Ohio

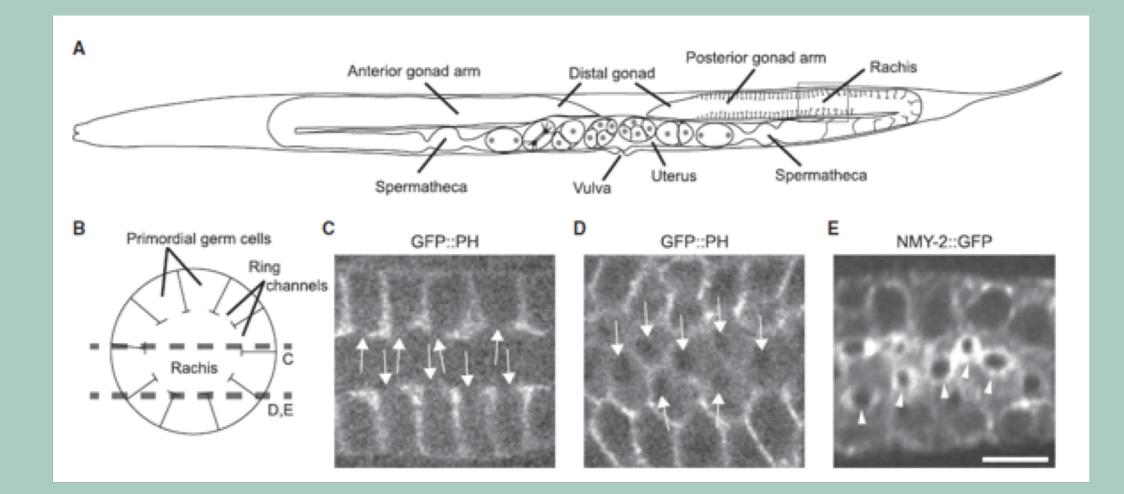
ABSTRACT Contractile rings play critical roles in a number of biological processes, including oogenesis, wound healing, and cytokinesis. In many cases, the activity of motor proteins such as nonmuscle myosins is required for appropriate constriction of these contractile rings. In the gonad of the nematode worm *Caenorhabditis elegans*, ring channels are a specialized form of contractile ring that are maintained at a constant diameter before oogenesis. We propose a model of ring channel maintenance that explicitly incorporates force generation by motor proteins that can act normally or tangentially to the ring channel opening. We find that both modes of force generation are needed to maintain the ring channels. We demonstrate experimentally that the type II myosins NMY-1 and NMY-2 antagonize each other in the ring channels by producing force in perpendicular directions: the experimental depletion of NMY-1/theoretical decrease in orthogonal force allows premature ring constriction and cellularization, whereas the experimental depletion of NMY-2/theoretical decrease in tangential force opens the ring channels and prevents cellularization. Together, our experimental and theoretical results show that both forces, mediated by NMY-1 and NMY-2, are crucial for maintaining the appropriate ring channel diameter and dynamics throughout the gonad.



Adriana Dawes, OSU

In the worm C. elegans: ring channels allow for nutrient exchange in development.

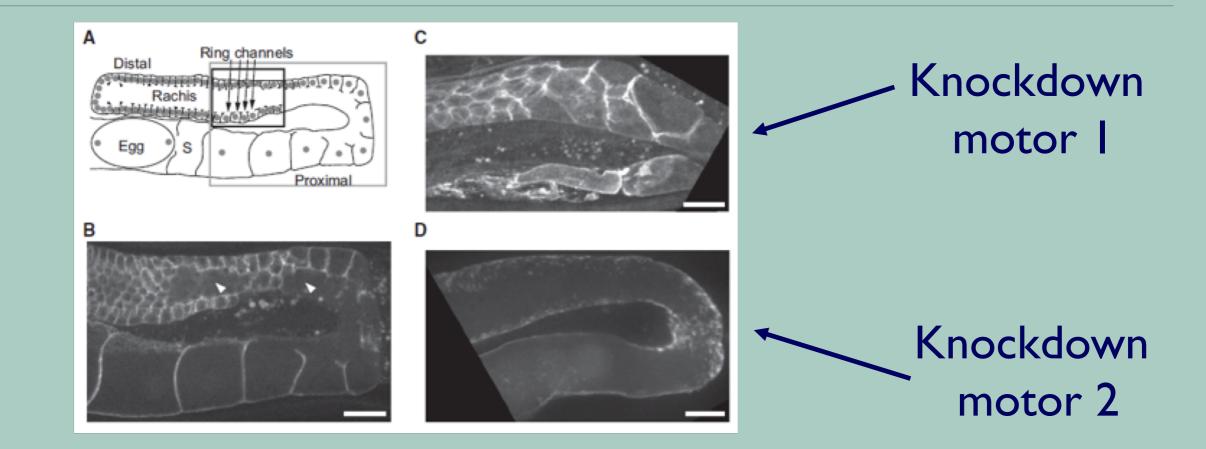
SPECIFIC MOTIVATION



□ Ring channels (formed of actin polymers) in the worm.

Coffman et al, 2016

THE QUESTION



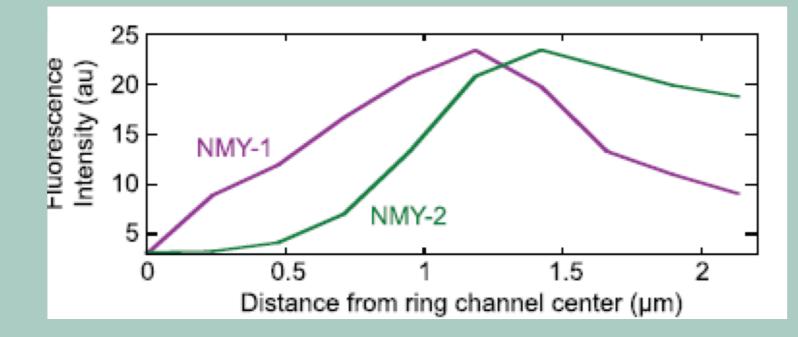
Two types of myosin motors, believed to function similarly, are involved in ring channel dynamics.

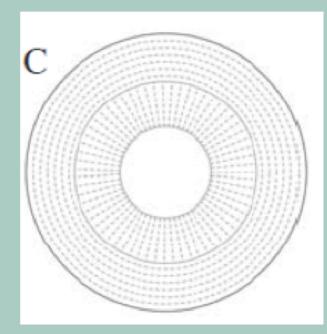
Experiments show that they antagonize each other with respect to cellularization.

Coffman et al, 2016

GOALS

Investigate ring formation using agent-based models for actin filaments and motors.
 Identify potential mechanistic differences between motors.





Coffman et al, 2016

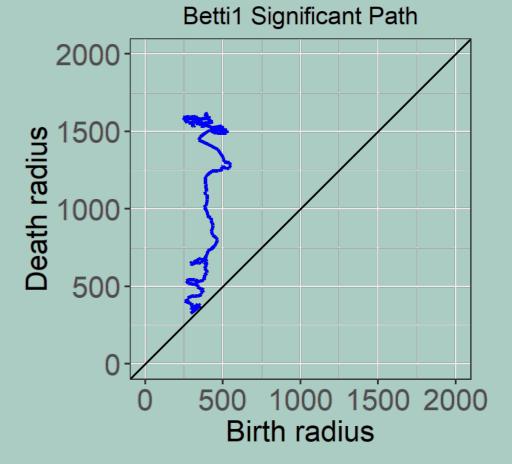
OUTLINE

I. Agent-based model simulations of actinmyosin interactions and data analysis measures

2. Detecting ring structure in time-series simulation data

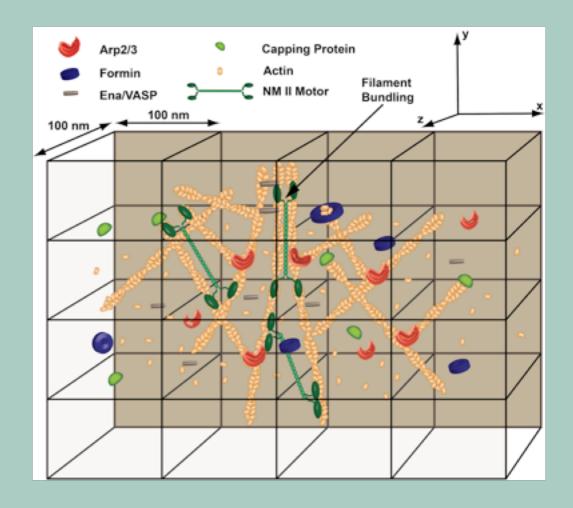
3. Connection with in vitro experimental data

10s



I. AGENT-BASED MODELING AND SIMULATION

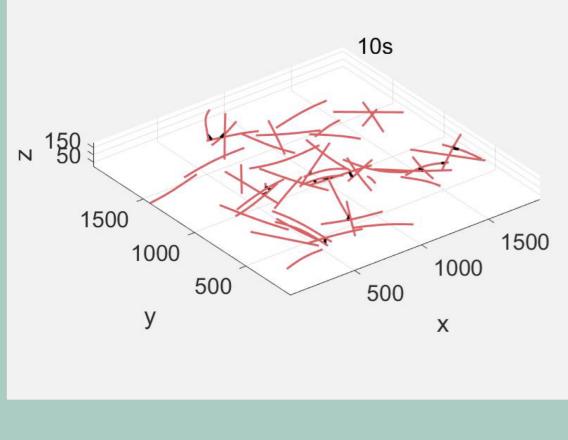
- Accounts for dynamics and molecular transport of chemical species.
- Diffusion and active transport
 are modeled as stochastic jumps
 between compartments.
- Is based on energy minimization.



Medyan (Papoian Lab)

Papoian Lab, University of Maryland; Popov, Komianos, and Papoian, 2016

DATA ANALYSIS MEASURES



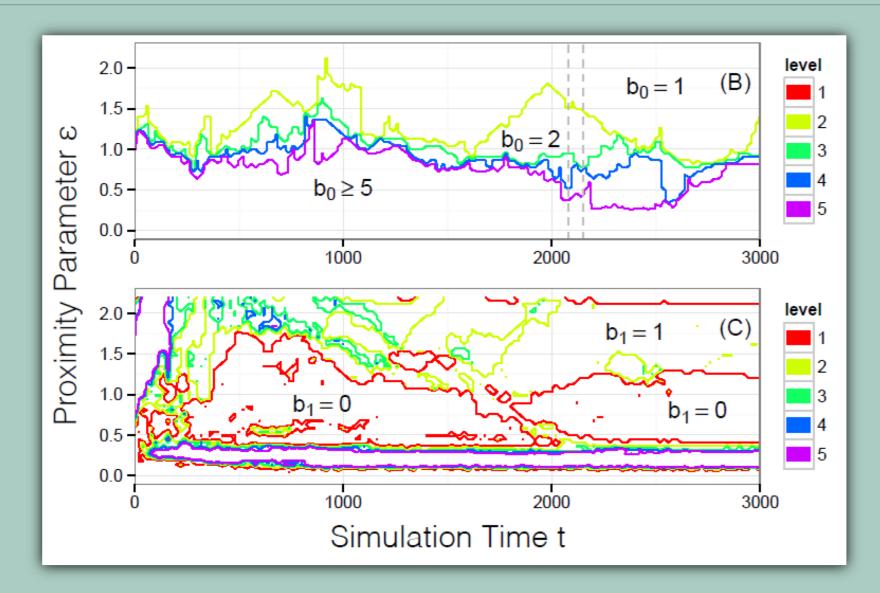




actin myosin crossfilament motor linker Riley Juenemann, Tulane University Scott McKinley, Tulane University

Contractility, alignment, filament length distributions

2. DETECTING RING STRUCTURE



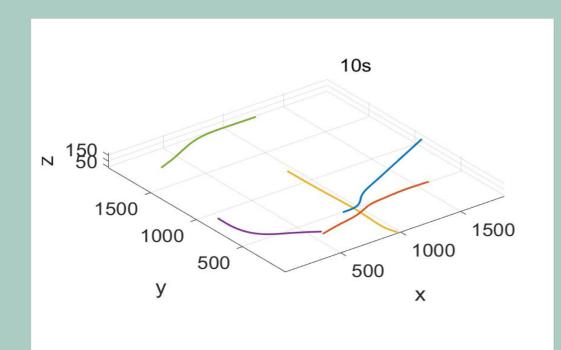


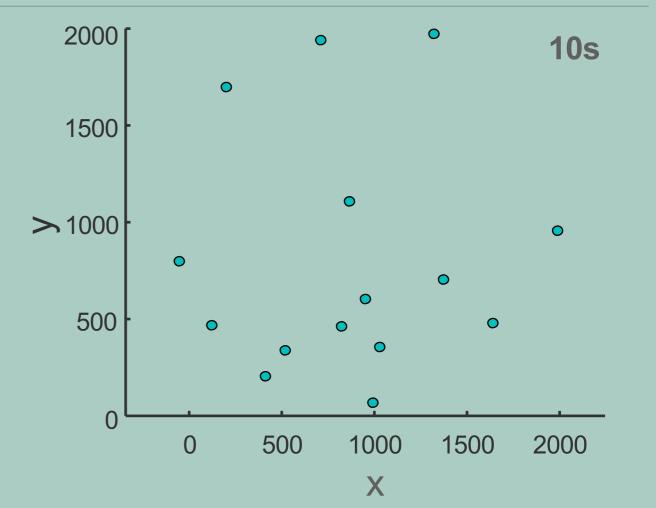
Chad Topaz, Williams College

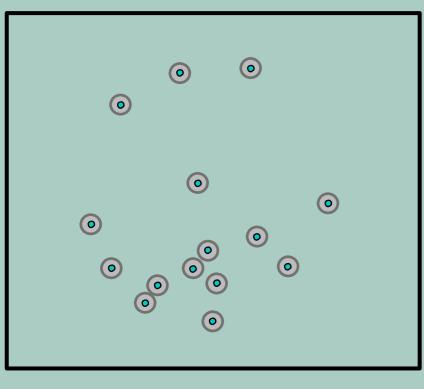
Inspiration: Crocker plots (Topaz, Zieglemeier, Halverson)

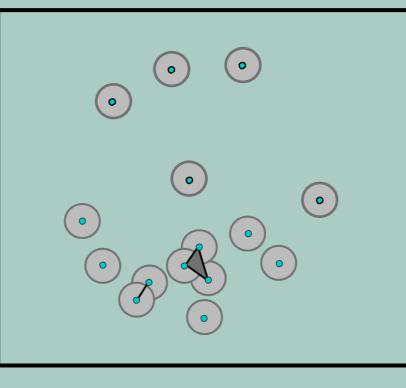
Topaz, Zieglemeier, Halverson, 2015

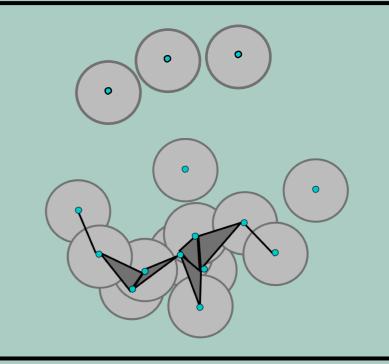
FRAMEWORK

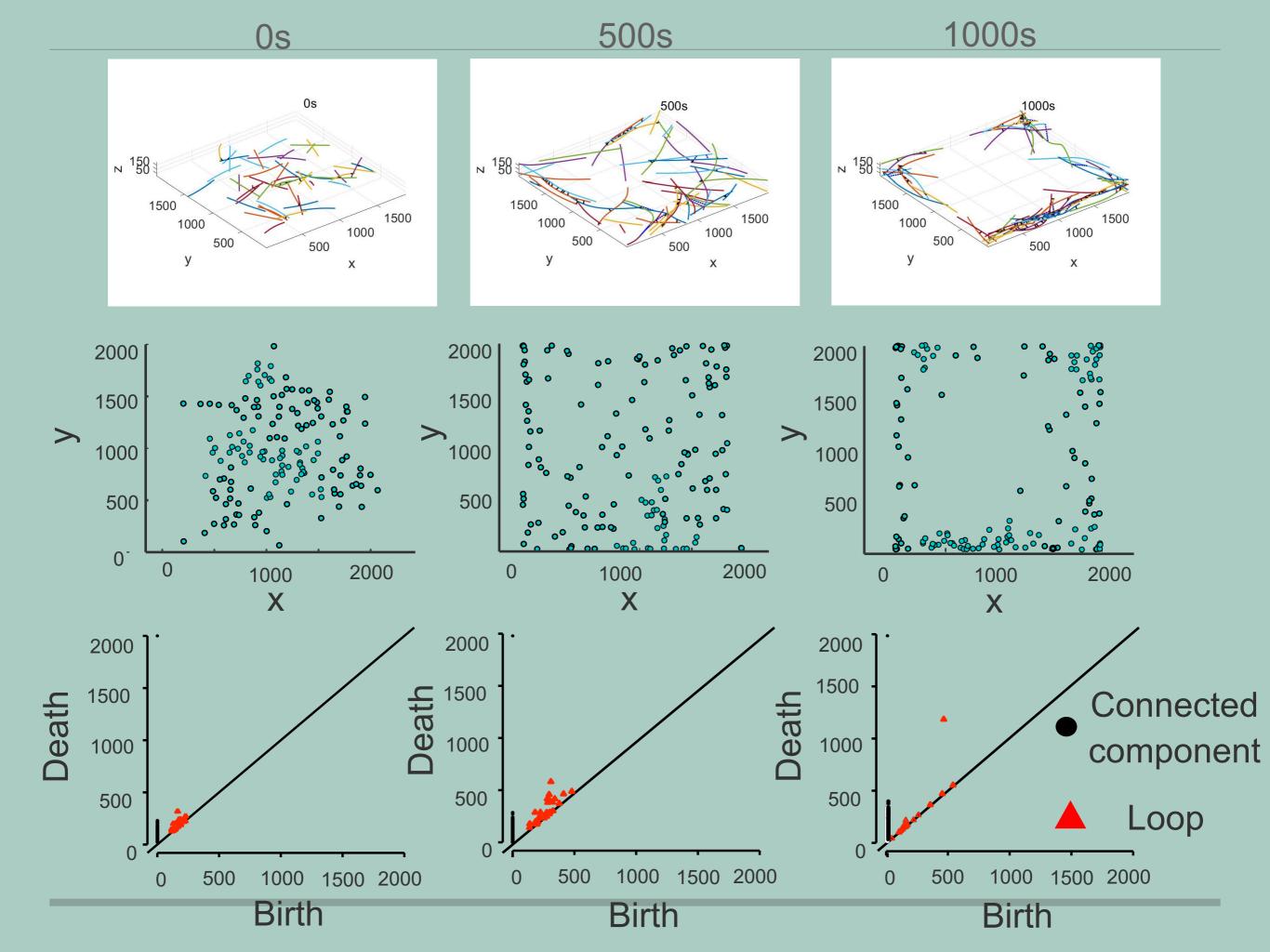


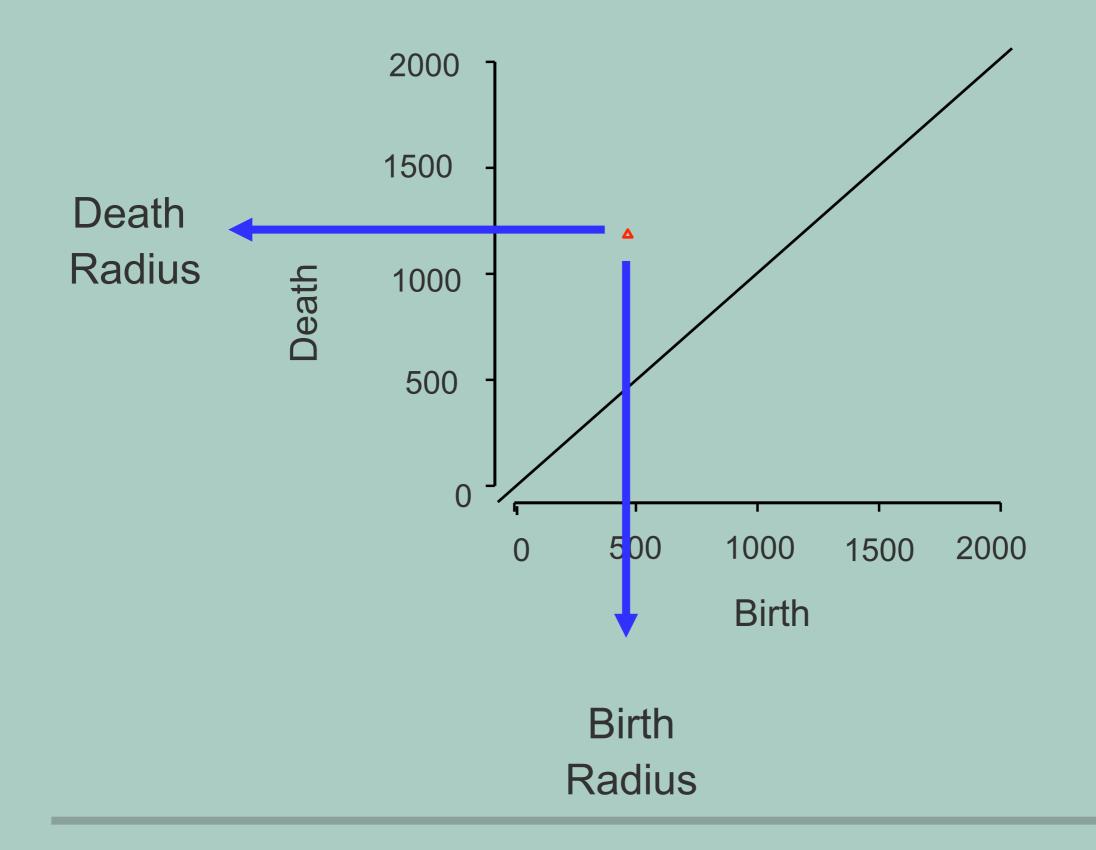


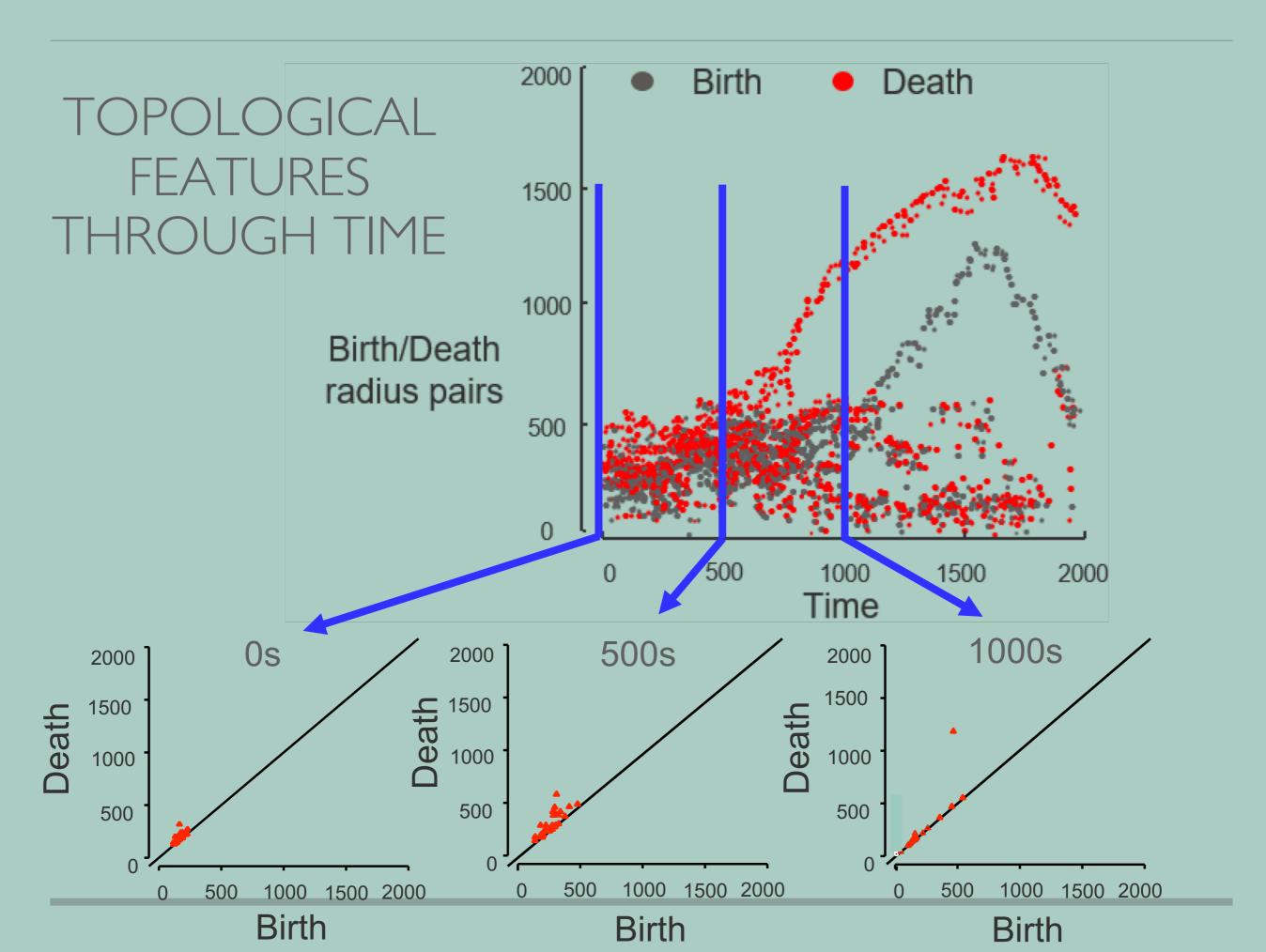




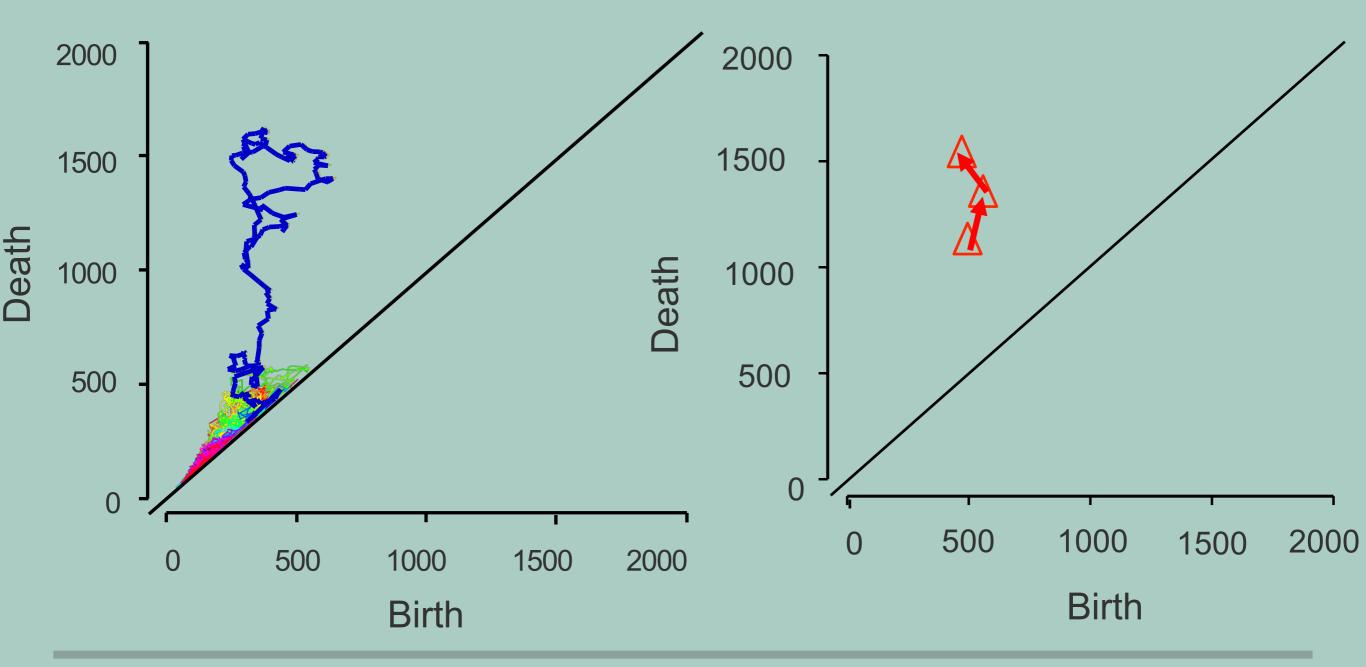






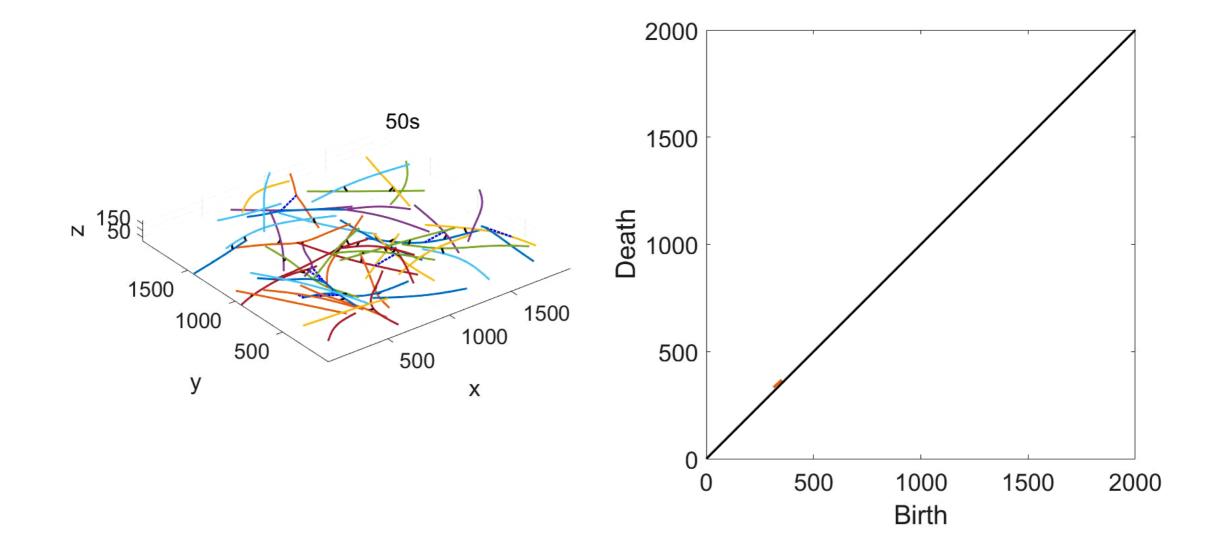


Propose a method for connecting pairs through time.
 Extract the most significant Betti I path.



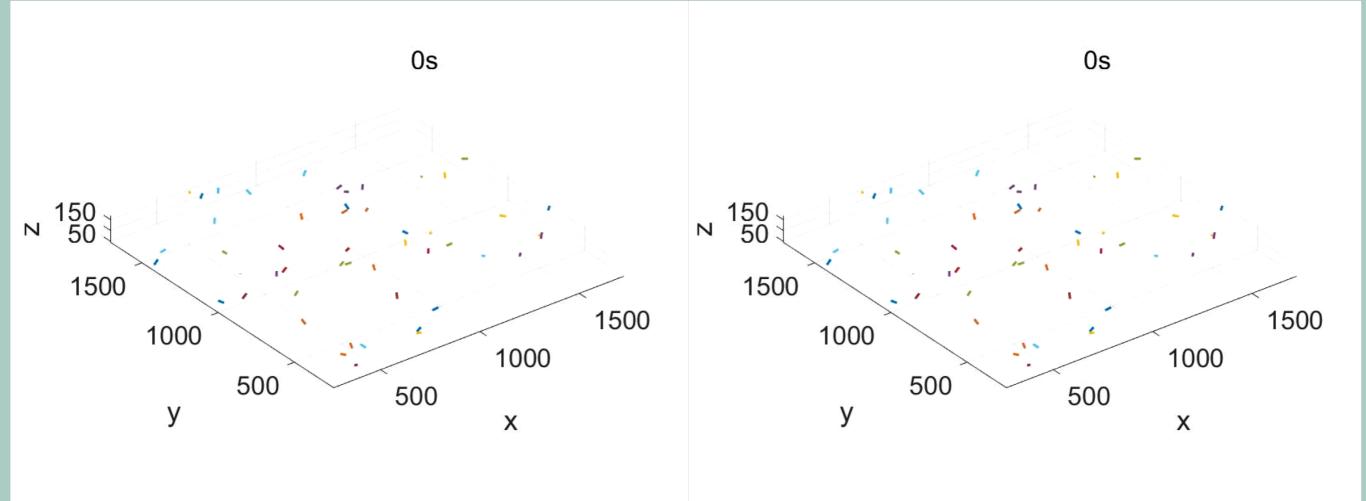
With Riley Juenemann and Scott McKinley (in preparation)

VISUALIZATION OF SIGNIFICANT PATH (RING STRUCTURE) EMERGENCE



With Riley Juenemann and Scott McKinley (in preparation)

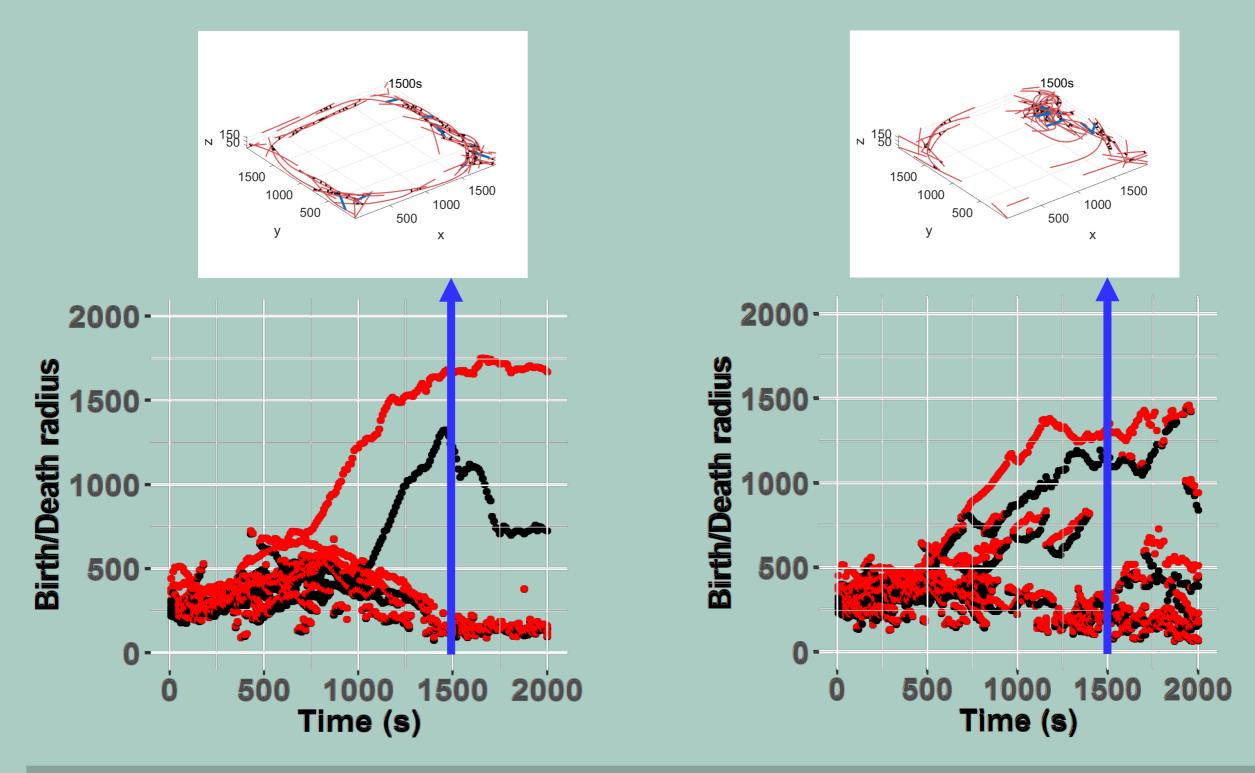
EXPLORE PARAMETER DIFFERENCES: ON-RATE



Small on-rate

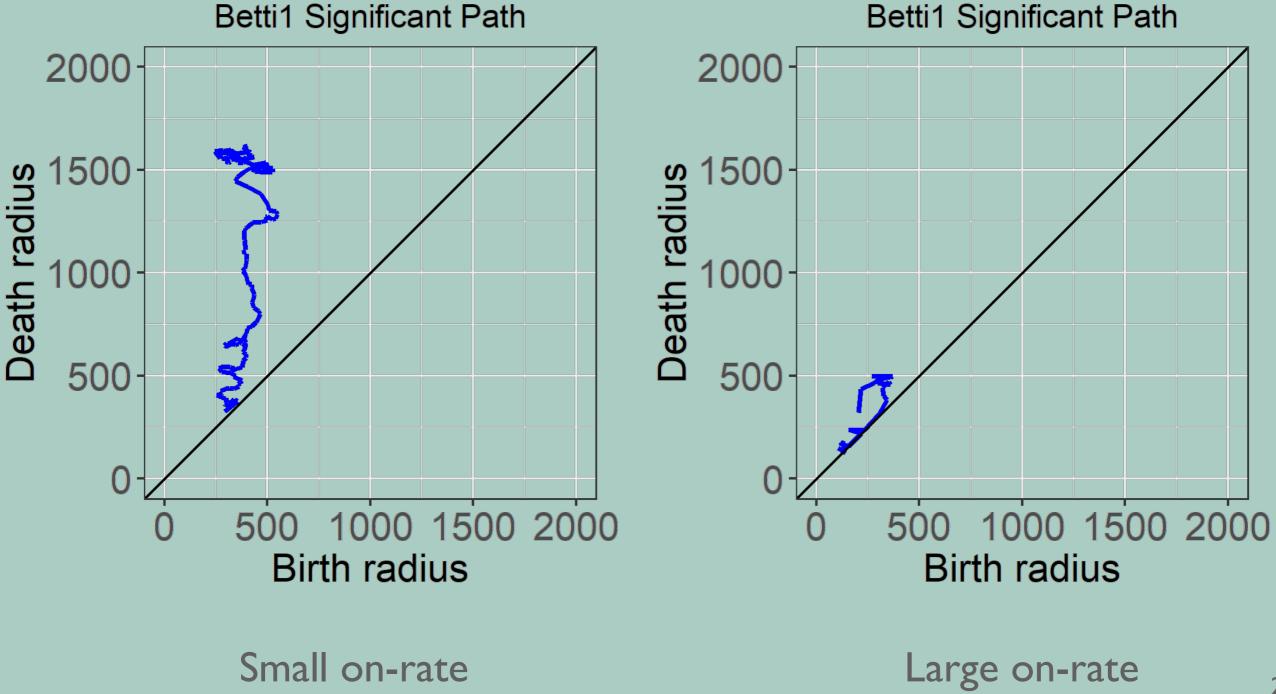
Large on-rate

TOPOLOGICAL FEATURES THROUGH TIME



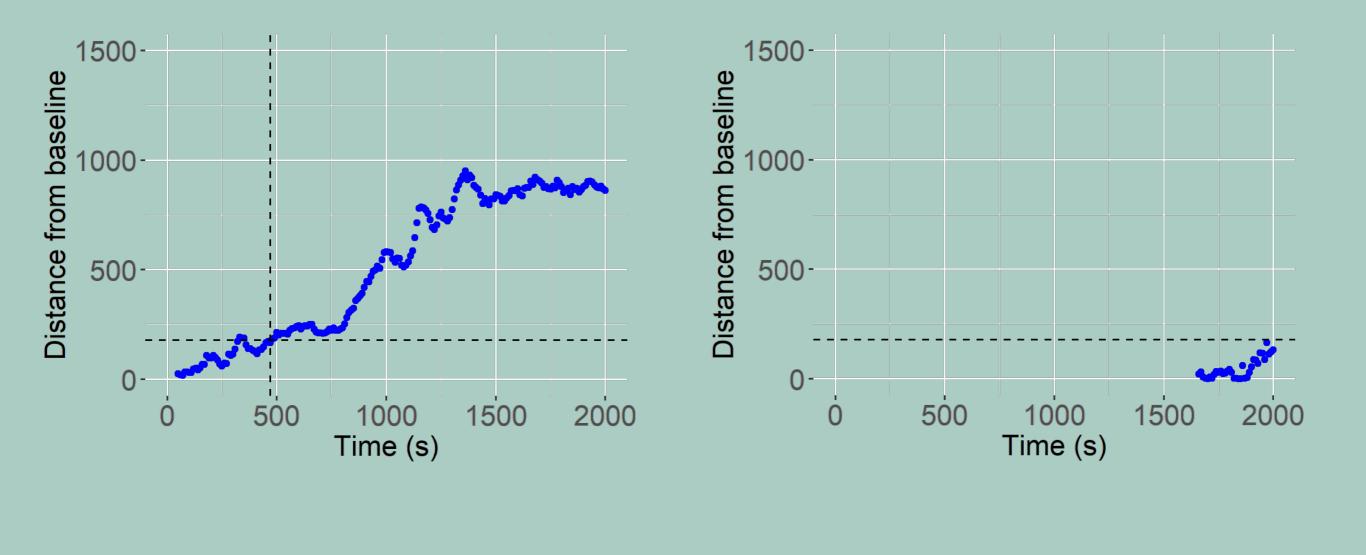
With Riley Juenemann and Scott McKinley (in preparation)

DETECTING RING STRUCTURE



With Riley Juenemann and Scott McKinley (in preparation)

DETECTING RING STRUCTURE

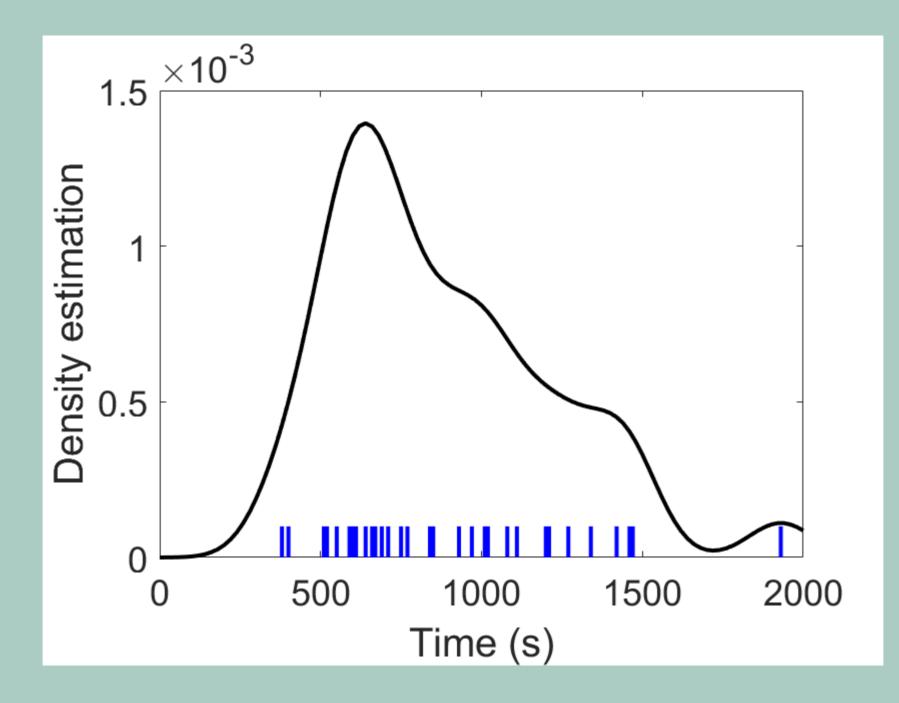


Small on-rate

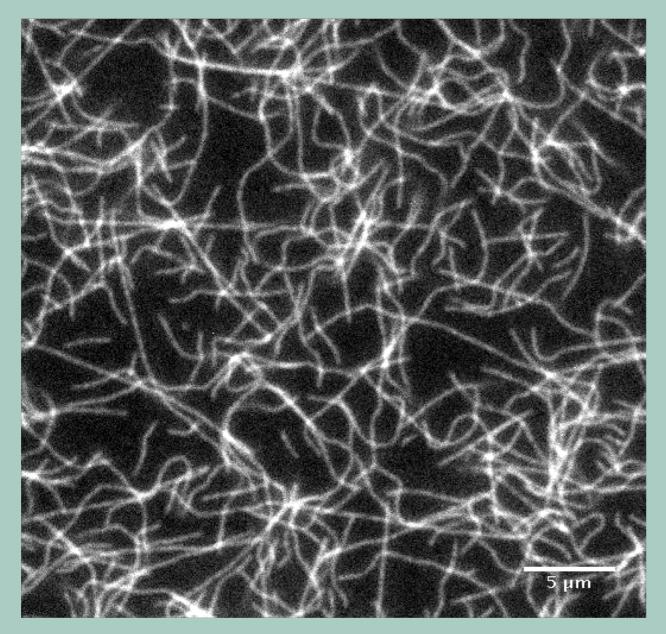
Large on-rate

With Riley Juenemann and Scott McKinley (in preparation)

TIME OF RING FORMATION



3. CONNECTION TO IN-VITRO EXPERIMENTAL DATA



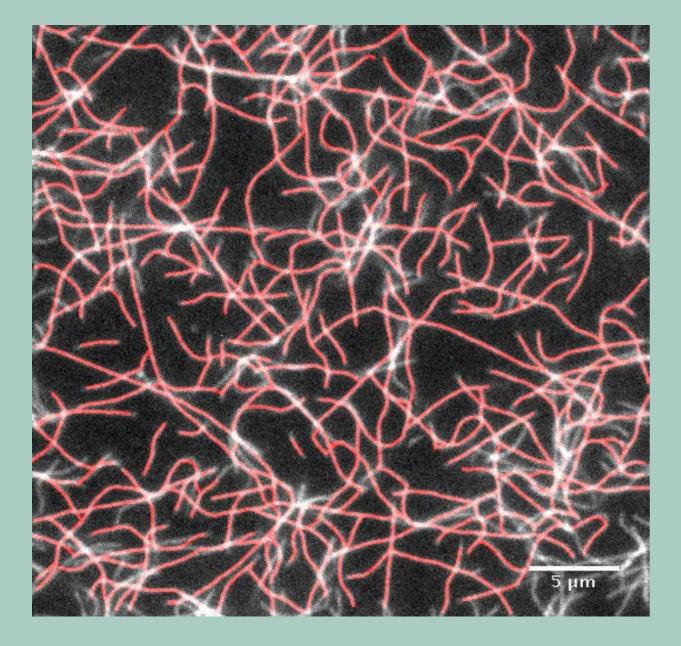
Myosin VI – actin interactions

Tim Atherton Tufts University

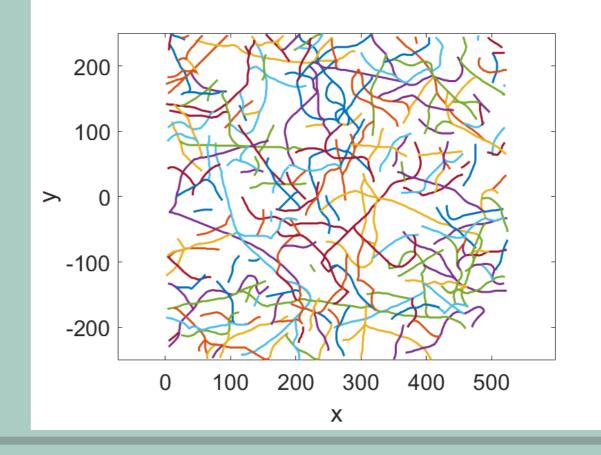


David Altman Willamette University

CONNECTION TO IN-VITRO EXPERIMENTAL DATA

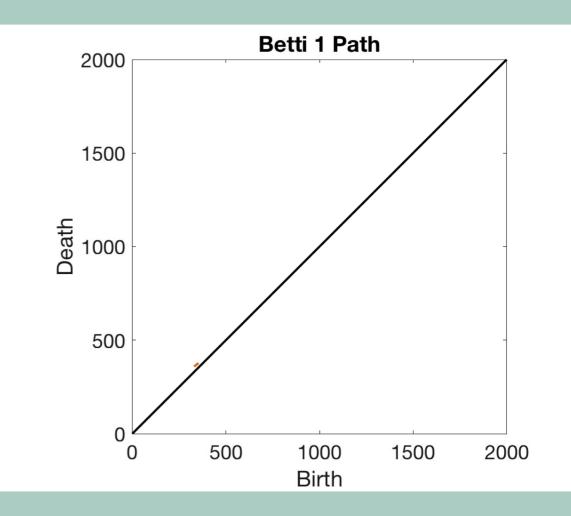


- Trace the actin filaments
- Apply data analysis measures and connect to simulations

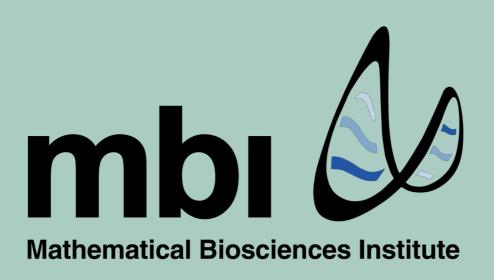


FUTURE WORK

- Modeling and simulation of ring formation and maintenance with realistic biological mechanisms
- Stability? Rigorous measure of significant paths?
- Comparison/incorporation into vines and vineyards¹ framework.



ACKNOWLEDGMENTS





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Thank you for your attention! go.osu.edu/veronicaciocanel $v_{ciocanel}$